Contents lists available at ScienceDirect



Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Syntheses of water-soluble [60]fullerene derivatives and their enhancing effect on neurite outgrowth in NGF-treated PC12 cells

Hiroki Tsumoto^a, Syo Kawahara^a, Yuki Fujisawa^a, Takayoshi Suzuki^a, Hidehiko Nakagawa^a, Kohfuku Kohda^{b,*}, Naoki Miyata^{a,*}

^a Graduate School of Pharmaceutical Sciences, Nagoya City University, 3-1 Tanabedori, Mizuho-ku, Nagoya, Aichi 467-8603, Japan ^b Graduate School of Pharmaceutical Sciences, Musashino University, 1-1-20 Shinmachi, Nishitokyo-shi, Tokyo 202-8585, Japan

ARTICLE INFO

Article history: Received 10 November 2009 Revised 21 January 2010 Accepted 25 January 2010 Available online 2 February 2010

Keywords: [60]Fullerene C₆₀ Water-soluble Neurite outgrowth PC12 cell

$A \hspace{0.1in} B \hspace{0.1in} S \hspace{0.1in} T \hspace{0.1in} R \hspace{0.1in} A \hspace{0.1in} C \hspace{0.1in} T$

Water-soluble [60]fullerene (C_{60}) derivatives were synthesized to examine their bioactivities. PC12 cells were used as a model of nerve cells and the bioactivities of synthesized C_{60} derivatives together with some reported ones were tested. Among the compounds tested, $C_{60}/(\gamma-CyD)_2$, C_{60} -bis(γ -CyD) (**5**) containing C_{60} -mono(γ -CyD) (**5**), and C_{60}/PVP were sufficiently soluble in water and showed an enhancing effect on the neurite outgrowth of NGF-treated PC12 cells.

© 2010 Elsevier Ltd. All rights reserved.

Since the discovery of [60] fullerene (C_{60}) by Kroto et al. in 1985,¹ extensive studies aimed at utilizing this molecule in different scientific fields have been carried out because of its unique physical and chemical properties (antioxidant, radical scavenger, photosensitizer, etc.). As regards the biological activities of fullerenes, antiviral,² antitumor,³ antimicrobial,⁴ and antioxidant activites,⁵ including inhibition of HIV protease,⁶ HIV reverse transcriptase,⁷ and DNA damage,⁸ have been reported. One drawback, however, is that the extremely low solubility of fullerenes in water hinders their application in biological systems. Various trials to make fullerenes water-soluble have been carried out. The tris-malonic acid derivative of C₆₀ (carboxyfullerene),⁹ polyhydroxylated C₆₀ (fullerol), C₆₀ wrapped with poly(vinylpyrrolidone) (C₆₀/PVP),¹⁰ and C₆₀ bicapped with γ -cyclodextrine (γ -CyD) $[C_{60}/(\gamma$ -CyD)₂]¹¹ have been demonstrated as water-soluble C₆₀ derivatives. Using C₆₀/PVP, we previously demonstrated its effect on promoting mouse embryonic limb bud cell differentiation¹² and its DNA cleaving activity.¹³

In this study, we synthesized other water-soluble C_{60} derivatives and examined their bioactivities as well as those of previously reported water-soluble C_{60} derivatives on nerve cells. As regards the bioactivity of fullerenes on nerve cells, carboxyfullerene is reported as a neuroprotective agent.¹⁴ In this Letter, we describe that water-soluble C_{60} derivatives are able to enhance neurite outgrowth in nerve growth factor (NGF)-treated PC12 cells that were used as a model of nerve cells.

The syntheses of water-soluble C_{60} derivatives **1**–**4** are shown in Scheme 1. The condensation reaction of compound 6^{15} with β -Dglucopyranosylamine 2,3,4,6-tetraacetate¹⁶ yielded C₆₀ derivative 1. Hydrolysis of compound 1 gave C₆₀ derivative 2. Activated ester 7^{17} was prepared by the condensation of compound **6** with *N*-hydroxysuccinimide (NHS). The reactions of compound **7** with mono(6-amino-6-deoxy)- γ -cyclodextrin (NH₂- γ -CyD)¹⁸ gave C₆₀ derivative 3 in 78% yield. Activated ester 9 of compound 8¹⁹ was prepared following the procedure for compound 7 and it was allowed to react with NH₂- γ -CyD to give C₆₀ derivative **4** in 80% yield. The synthesis of water-soluble C_{60} derivative **5** is shown in Scheme 2. C_{60} derivative **10**²⁰ was prepared by use of the Bingel reaction where 1,3-bis[2-(1,1-dimethylethoxy)-2-oxoethyl]propanedioate and C₆₀ were allowed to react with I₂ and DBU in dry toluene. Removal of the t-butyl groups of compound 10 gave compound 11. The condensation reaction of compound 11 with NHS gave activated ester 12. The reaction of compound 12 with NH₂- γ -CyD gave a mixture of C₆₀-bis(γ -CyD) (**5**) and C₆₀-mono(γ -CyD) (5'), the existence of which was confirmed by MALDI mass spectroscopy. The C_{60} -bis(γ -CyD)/ C_{60} -mono(γ -CyD) ratio was estimated to be almost 1/1 on the basis of MALDI signal intensities. Without further separation, C_{60} -bis(γ -CyD) (5) containing C_{60} -mono(γ -CyD) (**5**') was used in the biological assay. For the synthesis of C_{60} -bis(γ -CyD) and C_{60} -mono(γ -CyD), activated ester **12**

^{*} Corresponding authors. Tel./fax: +81 42 468 9204 (K.K.); tel./fax: +81 52 836 3407 (N.M.).

E-mail addresses: kohda@musashino-u.ac.jp (K. Kohda), miyata-n@phar.nagoya-cu.ac.jp (N. Miyata).



Scheme 1. Syntheses of C₆₀ derivatives 1–4. Reagents, conditions, and yields: (a) β-D-glucopyranosylamine 2,3,4,6-tetraacetate, EDCI, HOBt, DMF, 27–45%; (b) K₂CO₃, MeOH/ CHCl₃, 76%; and (c) NH₂-γ-CyD, DMF, 78–80%.



Scheme 2. Synthesis of C₆₀-bis(γ -CyD) (5) containing C₆₀-mono(γ -CyD) (5'). Reagents, conditions, and yields: (a) *p*-TsOH, toluene, reflux, 96%; (b)NHS, EDCI, DMF; and (c) NH₂- γ -CyD, DMF, 19% from 11.

was used instead of compound **11** to introduce the NH₂- γ -CyD moiety. By using the activated ester, purification of C₆₀-bis(γ -CyD) and C₆₀-mono(γ -CyD) was facilitated because the separation of unreacted compound **11** was extremely difficult. Details of the synthetic procedures and the characterization of new compounds **1–4**, **11**, and **12** are available in Supplementary data.

 $C_{60}/(\gamma$ -CyD)₂ is reported to form a complex in which two γ -CyDs are capping the C_{60} molecule.¹¹ The ¹H NMR spectrum of $C_{60}/(\gamma$ -CyD)₂ showed that the signals of the γ -CyD unit shifted slightly up- or downfield compared to those of γ -CyD itself. The UV spectrum of $C_{60}/(\gamma$ -CyD)₂ in aqueous solution showed a sharp absorption peak of C_{60} appearing at approximately 330 nm.²¹ In contrast, in the ¹H NMR spectrum of C_{60} -bis(γ -CyD), the same chemical shifts of the γ -CyD unit as those of γ -CyD itself were noted. Further, the UV spectrum of C_{60} -bis(γ -CyD) in aqueous solution showed a broad peak indicating that C_{60} -bis(γ -CyD) forms aggregates in aqueous solution. From these results, together with the results of a computer-based model study (data not shown), the γ -CyD moieties of C_{60} -bis(γ -CyD) may not form complex structures similar to those of $C_{60}/(\gamma$ -CyD)₂.

The antioxidant activity of compounds **1–4** and C₆₀-bis(γ -CyD) (**5**) containing C₆₀-mono(γ -CyD) (**5**') was examined using ESR spectrometry. 'OH was generated by use of the H₂O₂–Cu²⁺ system in the absence or presence of C₆₀ derivatives, and the generated 'OH was trapped by 5,5-dimethyl-1-pyrroline-N-oxide (DMPO). Then, the ESR spectra of 'OH-trapped DMPO (DMPO-OH) were measured. All C₆₀ derivatives examined reduced the signal intensity of DMPO-OH, indicating that these derivatives have antioxidative activity (Supplementary data).

The solubility in water of the fullerenes synthesized in this study was compared with typical water-soluble fullerenes, $C_{60}/(\gamma$ -CyD)₂, C_{60}/PVP , and fullerol, which were prepared according to reported procedures or purchased. The most soluble fullerene was $C_{60}/(\gamma$ -CyD)₂ (more than 100 µM), followed by C_{60}/PVP , C_{60} -bis(γ -CyD) (**5**) containing C_{60} -mono(γ -CyD) (**5**'), compound **3**, and fullerol (less than 1 µM). As the solubility in water of the other compounds synthesized in this study was much lower than 1 µM, only C_{60} -bis(γ -CyD) (**5**) containing C_{60} -mono(γ -CyD) (**5**') and compound **3** were used together with $C_{60}/(\gamma$ -CyD)₂, C_{60}/PVP , and fuller-ol for the biological assay using PC12 cells.

Rat pheochromocytoma PC12 cells were used as a model of nerve cells. Treatment of PC12 cells with NGF resulted in the induction of differentiation and the formation of neurites. First, we examined whether or not C_{60} derivatives have similar activity to NGF. Experiments were performed using the most water-soluble C_{60} , $C_{60}/(\gamma$ -CyD)₂. However, no effects as seen with NGF were observed for $C_{60}/(\gamma$ -CyD)₂ up to the examined concentration of 100 μ M. Next, the effect of C₆₀ derivatives on neurite outgrowth in NGF-treated PC12 cells was examined. The experimental protocol is as follows: PC12 cells were seeded and cultured for 2 days in the medium with serum, and then the medium was changed to a fresh one without serum. NGF (50 ng/mL) and appropriate concentrations of fullerenes were added. After incubating for 3 days, the number of neurites was counted and the length of the outgrowth was measured. Figure 1 shows PC12 cells that were treated with NGF plus 0–50 μ M C₆₀/(γ -CyD)₂. NGF-treated PC12 cells formed neurites and both neurite number and length of outgrowth increased in a dose-dependent manner. The length of outgrowth, which was expressed as the number of times the cell body diameter, and the ratio of neurite cells to total cells are shown in Figure 2. The total number of neurites increased in a dose-dependent



Figure 2. Ratio of neurites to total number of cells when treated with $C_{60}/(\gamma$ -CyD)₂. The length of neurite outgrowth was expressed as the number of times the cell body diameter. Data are expressed as means ± S.D.



Figure 1. Neurite outgrowth enhancing effect of $C_{60}/(\gamma$ -CyD)₂. NGF 50 ng/mL plus $C_{60}/(\gamma$ -CyD)₂: (a) 0 μ M, (b) 12.5 μ M, (c) 25 μ M, and (d) 50 μ M.

manner, reaching a maximum at 25 μ M. At higher concentrations of 50 and 100 μ M, a decrease in the total number of neurites was observed. The reason is not clear at present, although we speculate that some toxic effects of the compounds may have caused the decrease. Free γ -CyD is present in C₆₀/(γ -CyD)₂ and its presumed maximum concentrations are 0.32–5.1 mM relative to the concentrations of C₆₀/(γ -CyD)₂ of 6.25–100 μ M. Then, the effect of γ -CyD itself on neurite outgrowth was examined, but no enhancing effect of γ -CyD was observed (Supplementary data). A similar experiment was performed with C₆₀/PVP. The ratio of C₆₀ to PVP was approximately 1–17.6, and the concentrations of C₆₀/PVP (12.5 μ M/2.2 mM, 25 μ M/4.4 mM, and 50 μ M/8.9 mM) were examined. While PVP itself (2.2, 4.4, and 8.9 mM) had no effect, C₆₀/PVP showed neurite outgrowth enhancing effect (Supplementary data).

 C_{60} -bis(γ -CyD) (**5**) containing C_{60} -mono(γ -CyD) (**5**') was dissolved in 1% DMSO and its activity was examined. The concentration of C₆₀-bis(γ -CyD) (**5**) containing C₆₀-mono(γ -CyD) (**5**') was determined based on the average molecular weight of the mixture. The results in Figure 3 indicate that the ratio of neurites to the total number of cells increased in a dose-dependent manner to reach a maximum at 25 µM. This concentration is the same as that of $C_{60}/(\gamma$ -CyD)₂. At 50 μ M, the ratio of neurites to the total number of cells decreased, as seen for $C_{60}/(\gamma$ -CyD)₂. At 100 μ M, precipitates appeared. In the case of compound 3 and fullerol, precipitation was noted at 5 μ M and 1 μ M, respectively, and neither of them showed any enhancing effects probably because of their low concentration in the medium. C_{60} -bis(γ -CyD) (**5**) containing C_{60} -mono(γ -CyD) (5') was the most water-soluble among the compounds synthesized and its solubility was sufficient for use in biological assays. However, the water solubility of C_{60} -bis(γ -CyD) (5) containing C_{60} -mono(γ -CyD) (**5**') was lower than that of $C_{60}/(\gamma$ -CyD)₂. Nevertheless, compounds 5 and 5' have the advantage of being not a mixture of γ -CyD, such as C₆₀/(γ -CyD)₂. In addition, further chemical modification of compounds 5 and 5' is possible. Therefore, compounds **5** and **5**' could be used to study the real effects of C_{60} .

The effects of pretreatment with $C_{60}/(\gamma$ -CyD)₂ and C_{60} -bis(γ -CyD) (**5**) containing C_{60} -mono(γ -CyD) (**5**'), which showed the most potent neurite outgrowth enhancing effect, prior to NGF treatment on neurite outgrowth were examined. PC12 cells were seeded and cultured for 1 day in the medium with serum, and then 25 μ M $C_{60}/(\gamma$ -CyD)₂ or C_{60} -bis(γ -CyD) (**5**) containing C_{60} -mono(γ -CyD) (**5**') was added and the cells were incubated for another 2 days. The medium was changed to a fresh one without serum and only NGF (50 ng/mL) was added. After incubating for 3 days, the number of neurites was counted and the length of outgrowth was measured. The results are shown in Figure 4. Pre-incubation had a



Figure 3. Ratio of neurites to total number of cells when treated with C_{60} -bis-(γ -CyD) (**5**) containing C_{60} -mono(γ -CyD) (**5**'). The length of neurite outgrowth was expressed as the number of times the cell body diameter. Data are expressed as means ± S.D.



Figure 4. Effect of pretreatment of $C_{60}/(\gamma$ -CyD)₂ or C_{60} -bis(γ -CyD) (**5**) containing C_{60} -mono(γ -CyD) (**5**') on neurite outgrowth. Ratio of neurites to total number of cells when pretreated for 2 days with 25 μ M $C_{60}/(\gamma$ -CyD)₂ or C_{60} -bis(γ -CyD) (**5**) containing C_{60} -mono(γ -CyD) (**5**') before NGF (50 ng/mL) treatment. The length of neurite outgrowth was expressed as the number of times the cell body diameter. Data are expressed as means ± S.D.

greater neurite outgrowth enhancing effect than co-incubation for both C_{60} derivatives.

The mechanisms of neurite outgrowth enhancement by C_{60} derivatives are obscure. Ibi et al. reported that reactive oxygen species were produced by NADPH oxidase when PC12 cells underwent differentiation into neurites by NGF treatment.²² Reactive oxygen species inhibit neurite growth. As C_{60} is a scavenger of reactive oxygen species, it is possible that the C_{60} derivatives scavenged neuro-degenerative reactive oxygen species and as a result, neurite outgrowth was enhanced. Pretreatment with C_{60} derivatives had a greater neurite outgrowth enhancing effect than co-treatment. However, it is not clear if the C_{60} derivatives remained on the cell membrane even after the cells were washed or if the pretreatment with C_{60} derivatives resulted in switching on some receptor of cell membrane leading to show the activity. Further study is in progress.

Acknowledgment

This work was supported in part by MEXT-HAITEKU (2008).

Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2010.01.142.

References and notes

- 1. Kroto, H. W.; Heath, J. R.; O'Brien, S. C.; Curl, R. F.; Smalley, R. E. Nature 1985, 318, 162.
- Schinazi, R. F.; Sijbesma, R.; Srdanov, G.; Hill, C. L.; Wudl, F. Antimicrob. Agents Chemother. 1993, 37, 1707.
- 3. Tabata, Y.; Murakami, Y.; Ikada, Y. Jpn. J. Cancer Res. **1997**, 88, 110.
- Tsao, N.; Luh, T.-Y.; Chou, C.-K.; Wu, J.-J.; Lin, Y.-S.; Lei, H.-Y. Antimicrob. Agents Chemother. 2001, 45, 1788.
 Horie, M.; Fukuhara, A.; Saito, Y.; Yoshida, Y.; Sato, H.; Ohi, H.; Obata, M.;
- Horle, W., Fukuliata, A., Salo, F., Ioshuda, F., Salo, H., Ola, H., Olata, M., Mikata, Y.; Yano, S.; Niki, E. Bioorg. Med. Chem. Lett. **2009**, *19*, 5902.
 Friedman, S. H.; Ganapathi, P. S.; Rubin, Y.; Kenyon, G. L. J. Med. Chem. **1998**, *41*,
- rheunan, S. H., Ganapathi, P. S., Rubhi, F., Kenyon, G. L.J. Med. Chem. 1996, 41, 2424.
 Mashino, T.: Shimotohno, K.: Ikegami, N.: Nishikawa, D.: Okuda, K.: Takahashi,
- Mashino, T.; Shimotohno, K.; Ikegami, N.; Nishikawa, D.; Okuda, K.; Takahashi, K.; Nakamura, S.; Mochizuki, M. Bioorg. Med. Chem. Lett. 2005, 15, 1107.
- Tokuyama, H.; Yamago, S.; Nakamura, E.; Siraki, T.; Sugiura, Y. J. Am. Chem. Soc. 1993, 115, 7918.
- 9. Lamparth, I.; Hirsch, A. J. Chem. Soc., Chem. Commun. 1994, 1727.
- Yamakoshi, Y.; Yagami, T.; Fukuhara, K.; Sueyoshi, S.; Miyata, N. J. Chem. Soc., Chem. Commun. 1994, 517.
- 11. Yoshida, Z.; Takekawa, H.; Takekuma, S.; Matsubara, Y. Angew. Chem., Int. Ed. Engl. **1994**, 33, 1597.
- 12. Tsuchiya, T.; Oguri, I.; Nakajima, Y.; Miyata, N. Fullerene Sci. Technol. 1996, 4, 986.
- 13. Yamakoshi, Y.; Sueyoshi, S.; Fukuhara, K.; Miyata, N.; Masumizu, T.; Kohno, M. *J. Am. Chem. Soc.* **1998**, *120*, 12363.

- Dugan, L. L.; Turetsky, D. M.; Du, C.; Lobner, D.; Wheeler, M.; Almli, C. R.; Shen, C. K.-F.; Luh, T.; Choi, D. W.; Lin, T. Proc. Natl. Acad. Sci. U.S.A. **1997**, 94, 9434.
 Tada, T.; Ishida, Y.; Saigo, K. J. Org. Chem. **2006**, 71, 1633.
- 16. Reyes, B. C.; Jose, F. M.; Juan Antonio, G. P. Carbohydr. Res. 1986, 154, 280.
- Tsumoto, H.; Takahashi, K.; Suzuki, T.; Nakagawa, H.; Kohda, K.; Miyata, N. Bioorg. Med. Chem. Lett. 2008, 18, 657.
 Veonique, B.; Raphael, D.; Vinh, T.; Claude, R. Eur. J. Org. Chem. 2003, 24, 4810.
- Hummelen, J. C.; Knight, B. W.; LePeq, F.; Wudl, F.; Yao, J.; Wilkins, C. L. J. Org. Chem. 1995, 60, 532.
 Salvatore, F.; Andre, R. Comp. Rend. Chim. 2003, 6, 83.

- Guldi, D. M.; Hungerbüler, H.; Asmus, K.-D. J. Phys. Chem. **1995**, *99*, 13487.
 Ibi, M.; Katsuyama, M.; Fan, C. Y.; Iwata, K.; Nishinaka, T.; Yokoyama, T.; Yabe-Nishimura, C. Free Radical Biol. Med. 2006, 40, 1785.