

Dialysis Protocol

<u>final conc.</u>			<u>Buffer I</u>	<u>Buffer II</u>	<u>Stock</u>
6	<i>M</i>	Urea	72g		
20	<i>mM</i>	HEPES pH 7.9	4mL	40 mL	1 <i>M</i>
300	<i>mM</i>	KCl	30mL	44.76 g	2 <i>M</i>
0.1	%	NP-40	2mL	20 mL	10 %
5	<i>mM</i>	MgCl ₂	1mL	10 mL	1 <i>M</i>
10	%	Glycerol	20mL	200 mL	100 %
1	<i>mM</i>	DTT	200μL	2 mL	1 <i>M</i>
0.5	<i>mM</i>	PMSF	1mL	10 mL	100 <i>mM</i>
10	μ <i>M</i>	ZnSO ₄	200μL	2 mL	10 <i>mM</i>
fill w/ H ₂ O			200mL	2 L	

NOTE: add the ZnSO₄ if dealing with a Zn-finger protein

Procedure:

1. cut desired length of dialysis tubing; take note of the MW cut-off of the tubing
2. Boil tubing for 2' in dH₂O to sterilize
3. Soak tubing in Buffer I
4. Clamp up one end, open the other end, add some Buffer I into tubing, check for leaks
5. Remove buffer and add the protein sol'n. leave some space at top and clamp up.
6. Mix gently and let spin by stir bar slowly for 2hrs in the cold room (4C).
7. Dialyze as follows:

<u>Step</u>	<u>Urea</u>		<u>Time</u>	<u>Total V</u>
A	6M	buffer I	2 hrs.	200mL
B	5M	add 35mL buffer II	2 hrs.	236mL
C	4M	add 50mL buffer II	2 hrs.	286mL
D	3M	add 75mL buffer II	2 hrs.	361mL
E	2M	add 120mL buffer II good O/N point	2 hrs.	481mL
F	1M	subtract 250mL, add 250mL of buffer II	2 hrs.	500mL
G	0.5M	subtract 250mL, add 250mL of buffer II	2 hrs.	500mL
H	0M	dump all out, add buffer II only	2 hrs.	500mL
I	0M	dump all out, add buffer II only	2 hrs.	500mL

8. Transfer tubing to buffer III and spin for 2hrs. @ 4C (cold room)

<u>Buffer III:</u>	20mM	HEPES pH7.9	20mL	1M
	100mM	NaCl	20mL	5M
	0.1%	NP-40	10mL	10%
	5mM	MgCl ₂	5mL	1M
	10%	Glycerol	100mL	100%
	1mM	DTT	1mL	1M
	0.5mM	PMSF	5mL	100mM
	10μ <i>M</i>	ZnSO ₄	1mL	10mM
	fill w/ H ₂ O to		1L	

9. Open tubing and transfer contents to 15mL conical vial
10. Freeze on dry ice or liquid nitrogen and store at -80C to concentrate later.