

## Preparation of target RNA for hybridization to arrays by *in vitro* transcription using T3, T7 or SP6 RNA polymerase

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### Reagents

- ◆ DNA template (at 0.5–1 µg/µl)
- ◆ T3, T7 or SP6 RNA polymerase (Promega)
- ◆ Transcription buffer (5 × transcription buffer from Promega): 200 mM Tris-HCl pH 7.9, 30 mM MgCl<sub>2</sub>, 10 mM spermidine, 50 mM NaCl
- ◆ RNase inhibitor (recombinant RNasin®; Promega)
- ◆ 100 mM DTT (Promega)
- ◆ [ $\alpha$ -<sup>32</sup>P]UTP (3000 Ci/mmol) or [ $\alpha$ -<sup>33</sup>P]UTP (2500 Ci/mmol) (Amersham)
- ◆ rNTPs (Pharmacia): ATP, GTP, and CTP stored as 10 mM solutions, and UTP as 250 mM in nuclease-free distilled water (Promega)

### Method

The reaction below is for 20 µl using labelled UTP.

- 1 Add the following components to a microcentrifuge tube at room temperature in this order:
  - 5 × transcription buffer 4 µl
  - 100 mM DTT 2 µl
  - RNasin® 20 U
  - 10 mM ATP, GTP, and CTP 1 µl each
  - 250 µM UTP 1 µl
  - Template DNA 2 µl
  - [ $\alpha$ -<sup>32</sup>P]UTP or [ $\alpha$ -<sup>33</sup>P]UTP 2 µl
  - T3, T7 or SP6 RNA polymerase 20 U
  - Total volume 20 µl

- 2 Mix and incubate at 37 °C for 1 h.
- 3 Remove 1 µl for quantitation (see [Quantitation of transcripts](#)).
- 4 Remove unincorporated label by Sephadex® G25 or G50 column chromatography.<sup>a</sup>
- 5 Save 1 µl of the purified transcript for quantitation in a microcentrifuge tube.
- 6 Check the integrity of the transcript by electrophoresis on a polyacrylamide or an agarose gel.

## Notes

- a Ready-to-use columns are available from several suppliers including Promega and Pharmacia.